

Occurrence of triterpenyl acetates in soil and their potential as chemotaxonomical markers of Asteraceae

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ABSTRACT

As a part of a wider study aimed at determining new molecular biomarkers in soils and sediments that could be specific of distinct vegetation types with respect to unravelling past changes in land use, we analysed the neutral lipid content of soil developed in the catchment of a small lake in central France. The ketone/acetate lipid fraction of soil under pasture or meadow contains a series of pentacyclic triterpenyl acetates of wide structural diversity, most being reported in soil for the first time. The restricted number of potential plant sources of triterpenyl acetates (some produced by a single species) underlines the potential of triterpenyl acetates as new biomarkers to track

past vegetation change in palaeoenvironmental studies, when found in natural archives such as soil, sediments or peat.

Keywords: Asteraceae, soil, biomarkers, pentacyclic triterpenyl acetates.

1. Introduction

Understanding ecosystem response under natural (climatic) and anthropogenic constraints is of crucial importance for anticipating the consequences of current global change (Dearing, 2006). Land use changes have direct consequences for the budget of greenhouse gases at the global scale. At a more local scale, it has been demonstrated that soil properties result from a long history of land use (Hurt et al., 2006). The reconstruction of recent land use change can be approached through historical archives. For example, the impacts of drastic land use change on biodiversity are particularly well documented in the middle mountains of the French Massif Central during the twentieth century (Chassagne, 1956; Antonetti, 2006).

With respect to more ancient times, the scarcity of reliable written sources necessitates an investigation of natural archives (lake sediments, peat, and speleothems) in order to unravel past natural and anthropogenic impacts on environments. These environmental records retain information about such interactions in a well constrained time frame. Deciphering such interactions involves reconstruction of ancient land use via multidisciplinary studies (e.g. palaeoecology, archaeology, sedimentology; e.g. Berglund, 1991) that provide different and complementary information.

While the potential of sediments as natural archives has largely been proved, soils, although fundamental substrates of human subsistence and direct support of

agricultural practices, have not received comparable interest. Soils are known to be disturbed in terms of physical properties, organic matter (OM) content and thereby of fertility when affected by a land use change (Rumpel et al., 2009). After effects of such conversions remain perceptible in terms of biodiversity, and chemical and structural soil properties, at least at the historical scale (Dupouey et al., 2002). However, only a few studies have focussed on the direct recording of land use at the molecular level (e.g. van Bergen et al., 1997; Bull et al., 1998). These studies have shown that the lipid content is directly linked to the current overlying vegetation, since the major part of the soil lipids originates from plants and is well preserved (e.g. Dinel et al., 1990; van Bergen et al., 1997; Bull et al., 1998). This makes them a reliable tool as molecular biomarkers, i.e. as compounds preserved intact or slightly modified in natural archives (and aerosols), the structure of which allows unequivocal identification of their biological source (e.g. van Bergen et al., 1997; Oros and Simoneit, 2001a,b; Simoneit, 2002, 2004; Killops and Killops, 2004). Such molecular markers benefit from an increasing interest for studies dealing with vegetation change or human activity (e.g. Hjulström and Isaksson, 2009).

Amongst these, pentacyclic triterpenes are of special interest for organic geochemists exploring the distributions of lipids in soil, sediments and peat as remnants of past vegetation. For example, those with a lupane, oleanane or ursane structure, as well as their diagenetic derivatives, in soil and sediments are classically used as tracers of angiosperms (Cranwell, 1984; Jacob et al., 2007). However, as a result of their wide structural (skeletal) diversity (oleanane, ursane, lupane, taraxerane...), potential isomerism and associated functional groups, several recent studies have shown that, in favourable cases, pentacyclic triterpenes can be discriminant for more constrained taxa. For example, Gramineae are the most common producers of pentacyclic triterpenes

bearing a MeO group at C-3 (Ohmoto et al., 1970; Jacob et al., 2005; Zocatelli et al., 2010). In particular, more specific relationships can be established, such as that allowing the linking of miliacin (olean-18-en-3 β -ol methyl ether) in sediments from Lake le Bourget (French Alps) to *Panicum miliaceum* (broomcorn millet) cultivated in the watershed (Jacob et al., 2008, 2009).

Here we report the presence of a series of pentacyclic triterpenyl acetates in a soil developed under pasture in the catchment of Lake Aydat (Massif Central, France). Following description of the different compounds that were analysed by way of gas chromatography-mass spectrometry (GC-MS), we discuss their potential biological sources and finally evaluate their potential for paleoenvironmental studies.

2. Setting, materials and methods

2.1. Location and general context

Lake Aydat is ca. 25 km SW of Clermont-Ferrand, in the volcanic French Massif Central. The small catchment of the lake (ca. 30 km²) is at an altitude ranging from 825 m (lake level) to ca. 1,300 m above sea level (a.s.l.). The tops of the volcanoes, the highest points, are covered by coniferous forests (mainly of *Picea* sp.), whereas the slopes are covered by pasture/grassland and, to a lesser extent, shrubs. The geology, relating mainly to a volcanic origin, led to the development of andisols as a result of rapid chemical erosion in the presence of OM (Jones et al., 2005). Typically, a homogeneous organo-mineral sandy-silty horizon A, developed above the C horizon. These slightly acidic soils are well drained and quite shallow (< 30 cm depth).

2.2. Materials and methods

A total of 35 soil samples were taken from the lake catchment in the autumn of 2008. They represent the diversity of land use, as described above, and were collected at different altitude and orientation, so that each condition prevailing in the catchment was taken into account. In the absence of recognizable layers, soils were sub-sampled in slices 2 cm thick. The 0-2 and 2-4 cm depth slices were analysed, in random order, since they were considered the most representative of the current vegetation cover.

Samples were dried at 40 °C for 48 h in an oven, crushed in a mortar and sieved at 2 mm. An aliquot (ca. 2 g) of each was extracted by way of automatic solvent extraction with a Dionex® ASE 200 using a mixture of CH₂Cl₂:MeOH (1:1). After removal of the solvent under N₂, the extract was separated into neutral, acidic and polar fractions on aminopropyl-bonded silica as described by Jacob et al. (2005). The neutral fraction was further separated into aliphatics, aromatics, ethers and esters, ketones and acetates, and alcohols by way of flash chromatography with a Pasteur pipette filled with activated silica (24 h at 120 °C, then deactivated with 5% H₂O) and using a sequence of solvents of increasing polarity. The fraction containing triterpenyl acetates and ketones was eluted with a mixture of hexane:EtOAc (9:1) and 5 α -cholestane was added prior to analysis via gas chromatography-mass spectrometry (GC-MS) with a TRACE-GCQ Polaris. The GC instrument was fitted with a Rtx-5MS column (30 m x 0.25 mm i.d., 0.25 μ m film thickness; 5m column guard). Its operating conditions were: 40 °C (1 min) to 120 °C at 30 °C min⁻¹, then from 120° to 300 °C (held 30 min) at 5 °C min⁻¹. Samples were injected in splitless mode, with the injector at 280 °C. He was the carrier gas in constant flow mode at 1 ml min⁻¹. The mass spectrometer was operated in the electron ionization (EI) mode at 70 eV and scanned from *m/z* 50 to 600. Component identification was based on comparison of retention times and mass spectra with

authentic standards. The concentrations of triterpenyl acetates were estimated by measuring peak areas in ion chromatograms. After calculating a correction factor between the peak area on the specific ion chromatogram and the peak area on the total ion current (TIC) chromatogram, the TIC area of triterpenyl acetates was measured relative to that of the standard (5 α -cholestane) and to the mass of sample extracted. The identification of all compounds, except isopichierenyl acetate, was confirmed using authentic standards.

3. Results

This study reports on the triterpenyl acetates in a single soil sample that showed the widest diversity among the 35 samples. Variations in the distributions, probably reflecting various biological inputs resulting from local conditions in the catchment, are not discussed here.

Fifteen triterpenyl acetates eluted in the 44-52 min t_R range under our GC conditions (Fig. 1). The structures discussed are displayed together with the mass spectra in Fig. 2.

The spectra of all triterpenyl acetates display ions m/z 468, 453, 393 408 ($[M^+]$, $[M^+-15]$, $[M^+-60]$, i.e. loss of the acetate group, and $[M^+-60-15]$, respectively), with an intensity that depends on the compound. Compounds 1, 2, 3 and 6 elute in the 44-47 min t_R range and are characterised by intense m/z 189, 203, 204 and 218 fragments attesting to a taraxerane, oleanane or ursane structure (Shiojima et al., 1992).

Considering the relative retention times, 2, 3 and 6 were identified as olean-13(18)-en-3 β -yl acetate (δ -amyrin acetate), olean-12-en-3 β -yl acetate (β -amyrin acetate) and urs-12-en-3 β -yl acetate (α -amyrin acetate) respectively, whereas the rather intense

fragments at m/z 269, 329 and 344 in the mass spectrum of compound 1 attest to a tarax-14-en-3 β -yl acetate (taraxeryl acetate) structure.

The spectra of 10 and 11 are also characterized by intense fragments at m/z 189 and 204. Their late elution time vs. 2 and 3, combined with the presence of an ion at m/z 249, allowed identification of 10 as taraxast-20-en-3 β -yl acetate (ψ -taraxasteryl acetate) and 11 as taraxast-20(30)-en-3 β -yl acetate (taraxasteryl acetate). This assignment was confirmed by comparison with authentic standards.

Intense fragments at m/z 259 and 274, present in the spectrum of 5, are diagnostic for D:B-friedo-oleanane structures (Shiojima et al., 1992), consistent with glutin-5-en-3 β -yl acetate (glutiny acetate).

The spectra of 4, 8, 9, 12, 13 and 15 are characterized by intense ion at m/z 229, 289, 241 and 301, with a minor contribution from ions at m/z 255 and 393 (Fig. 2). These features are characteristic for D:C-friedo-ursen-3 β -ol and D:C-friedo-oleanen-3 β -ol acetates (bauerenyl and multiflorenyl acetates, respectively) and are also found in swertenyl acetates (Shiojima et al., 1992). The use of authentic standards and relative retention times allowed identification of 4 as bauer-8-en-3 β -yl acetate (isobauerenyl acetate), 9 as bauer-7-en-3 β -ol acetate (bauerenyl acetate), 8 as multiflor-7-en-3 β -yl acetate (multiflorenyl acetate), 12 as pichier-8-en-3 β -yl acetate (isopichierenyl acetate – no standard available), 13 as pichier-9(11)-en-3 β -yl acetate (pichierenyl acetate) and 15 as pichier-7-en-3 β -yl acetate (swertenyl acetate). Finally, 7 was identified as lup-20(29)-en-3 β -yl acetate (lupeyl acetate) and 14 as gammacer-16-en-3 β -yl acetate (gammacerenyl acetate) on the basis of their mass spectra (Shiojima et al., 1992) and authentic standards.

Taraxeryl acetate (1.10 $\mu\text{g/g}$ soil), pichierenyl acetate (0.91 $\mu\text{g/g}$ soil), glutinyl acetate (0.81 $\mu\text{g/g}$ soil), taraxasteryl acetate (0.29 $\mu\text{g/g}$ soil), ψ -taraxasteryl acetate (0.07 $\mu\text{g/g}$ soil) and isopichierenyl acetate (0.54 $\mu\text{g/g}$ soil) were the most abundant compounds in the sample. In all the other samples, concentrations rarely reach 1 $\mu\text{g/g}$ soil. For comparison, the concentrations in plants range from a few tens of μg to a few mg per gram dried plant, with a strong variability as to whether roots or aerial parts are considered (i.e. Bohlmann et al., 1981; Shiojima et al., 1989b; Lu et al., 1994; Tsao et al., 2008).

4. Discussion

4.1. Distribution of triterpenyl acetates in plants

Our literature survey of triterpenyl acetates sources led to an inventory of > 460 plants known to produce these compounds. Most information concerning Asteraceae (432 species among the 460) were extracted from the “Bohlmann Files” database (Jakupovic et al., 2011), which concerns all natural components of this family. This information was completed with other data on the distribution of triterpenyl acetates in Asteraceae. Additional data on the occurrence of the compounds in other plant taxa allowed testing of the chemotaxonomical value of the different triterpenyl acetates. Depending on bibliographical source, the degree of detail is highly variable for the different taxa. The resulting database evidently suffers from bias linked to a more specific interest toward a given plant taxa or a given chemical family. Nevertheless, the inventory provides key information on the distribution of pentacyclic triterpenyl acetates in plants.

The most commonly reported triterpenyl acetates are lupeyl acetates, comprising lup-20(29)-en-3 β -yl acetate, lup-19(20)-enyl acetate, tarolupenyl acetate (lup-19(21)-en-3 β -yl acetate), epilupeyl acetate (epi-lup-20(29)-en-3 β -yl acetate), lactucenyl acetate (D-friedo-lup-14-en-3 β -yl acetate; Shinozaki et al., 2011) and neolupenyl acetate (lup-12-en-3 β -yl acetate), the first being by far the most reported. Of the Asteraceae, 416 species are reputed to contain lupeyl acetates (Jakupovic et al., 2011; Madrigal et al., 1975), but this component is also present in various other families such as Amaranthaceae, Cucurbitaceae, Euphorbiaceae, Moraceae and Apocynaceae (e.g. Jewers and Manchada, 1970; Dutta and Ray, 1972; Woldu et al., 1988; Yoshizumi et al., 1998; Medeiros et al., 2001; Gasparetto et al., 2010). Taraxasteryl and ψ -taraxasteryl acetates were overwhelmingly detected in Asteraceae, with 153 and 29 occurrences, respectively (Dutta and Ray, 1972; Madrigal et al., 1975; Jordon-Thaden and Louda, 2003; Jakupovic et al., 2011), and also in a few Apocynaceae (6 and 2 occurrences, respectively; Jewers and Manchada, 1970; Thakur et al., 1984; Abbott et al., 1990; Sen et al., 1992; Medeiros et al., 2001; Zhang et al., 2006; Yin et al., 2007; Pereira et al., 2008). Bauerenyl acetate was found in three Asteraceae spp., four Apocynaceae and *Euphorbia chrysocoma* (Cava et al., 1967; Cicció-Alberti and Hoet, 1981; Lao et al., 1983; Shi et al., 2005; Pereira et al., 2008; Jakupovic et al., 2011). Isobauerenyl acetate was only found in *Euphorbia fischeriana* (Liu et al., 2001) and in two Asteraceae spp.: *Picris hieracioides* (Shiojima et al., 1989a) and *Centaurea aspera* (Picher et al., 1985); δ -amyrin acetate occurs in three Asteraceae spp.: *Vernonia cinerea* (Misra et al., 1984), *Inula britannica* and *Echinops echinatus* (Jakupovic et al., 2011) and in the Phyllanthaceae sp., *Phyllanthus polyanthus* (Ndlebe et al., 2008).

The synthesis of lupeyl, taraxasteryl, ψ -taraxasteryl, bauerenyl, as well as that of isobauerenyl acetates therefore appears almost exclusively constrained to Asteraceae. The lack of data concerning δ -amyrin acetate does not allow discussion of its specificity. Remarkable is the detection of pichierenyl, isopichierenyl, gammacer-16-en-3 β -yl and swertenyl acetates in our soil samples since these compounds have only been described in the roots of the hawkweed oxtongue *Picris hieracioides* (Shiojima et al., 1989a,b; 1995).

Although taraxeryl and β -amyrin acetates are also found in Asteraceae (9 and 29 spp., respectively; Yang et al., 1994; Madrigal et al., 1975; Jakupovic et al., 2011), they were also detected in various taxa such as Apocynaceae, Aceraceae, Euphorbiaceae, Crassulaceae, Betulaceae and Moraceae (e.g. Dutta and Ray, 1972; Stevens et al., 1994a; b; van Bergen et al., 1997; Vilegas et al., 1997; Wada et al., 1998; Pereira et al., 2008). Multiflorenyl acetate was found in *Cirsium* sp. and *Ixeris chinensis* (Asteraceae; Ulubelen and Berkan, 1977; Shiojima et al., 1994; Kataria, 1995; Jakupovic et al., 2011), as well as in *Tabernaemontana longipes* (Apocynaceae; Cicció-Alberti and Hoet, 1981), *Euphorbia guyoniana* (Euphorbiaceae; Haba et al., 2007), *Sedum brevifolium* and *S. meyeri-johannis* (Stevens et al., 1994a), and in *Polypodium niponicum* (Polypodiaceae; Ageta and Arai, 1983). α -Amyrin acetate was described in 11 Asteraceae spp. (Jakupovic et al., 2011), but also in various families such as Apocynaceae spp. [e.g. *Tabernaemontana laeta* (Medeiros et al., 2001), Crassulaceae (Stevens et al., 1994a), Sapotaceae (e.g. *Pouteria tomentosa*; Anjaneyulu, 1965), Moraceae (e.g. *Antiaris africana*; Okogun et al., 1976), or Balanophoraceae (*Balanophora abbreviate*; Yadagiri et al., 1984)]. Therefore, taraxeryl, β -amyrin, α -amyrin and multiflorenyl acetates cannot be considered as being specific of Asteraceae.

Finally, glutinyl acetate has not been reported in Asteraceae but has been in *Kalanchoe daigremontiana* (Crassulaceae; van Maarseveen and Jetter, 2009), *Acer manschuricum* (Aceraceae; Ding et al., 2010), *Erythrophleum fordii* (Fabaceae; Tsao et al., 2008), as well as in *Dorstenia* sp. and *Maquira coriacea* (Moraceae; Woldu et al., 1988; Mitaine-Offer et al., 2001).

4.2. Comparison of soil triterpenyl acetate diversity with local plant sources

According to the survey of Antonetti et al. (2006), 97 species of Asteraceae occur in the catchment of Lake Aydat, ten being reputed to produce triterpenyl acetates (Table 2): the greater burdock (*Arctium lappa*), the musk thistle (*Carduus nutans*), the creeping thistle (*Cirsium arvense*), the mouse-ear hawkweed (*Hieracium pilosella*), the cotton thistle (*Onopordum acanthium*), the bristly oxtongue (*Picris echioides*), the hawkweed oxtongue (*Picris hieracioides*), the goldenrod (*Solidago virgaurea*), the common sowthistle (*Sonchus oleraceus*) and the dandelion (*Taraxacum* sp.).

Picris hieracioides, largely present in the area, appears as the most evident primary source of triterpenyl acetates in our samples because it is the only plant reputed to produce isopichierenyl, pichierenyl, swertenyl and gammacer-16-en-3 β -yl acetates (Shiojima et al., 1989a; b). Because it also produces α -amyrin, β -amyrin, lupeyl, bauerenyl, isobauerenyl, taraxasteryl and ψ -taraxasteryl acetates (Shiojima et al., 1989a; b; 1995), this plant could also have contributed to these compounds in soil.

Other local sources of β -amyrin acetate from Asteraceae can alternatively be provided by *Cirsium arvense* (Tulloch and Hoffman, 1982) and *Solidago virgaurea* (Choi et al., 2004). Similarly, taraxasteryl acetate can be produced by up to seven species of Asteraceae encountered in the catchment (*Arctium lappa*, *Carduus nutans*,

Cirsium arvense, *Onopordum acanthium*, *Sonchus oleraceus* and *Taraxacum officinale*; Jakupovic et al., 2011; Jordon-Thaden and Louda, 2003; Dutta and Ray, 1972; Khalilova et al., 2004; Hänsel et al., 1980). ψ -Taraxasteryl acetate is also synthesised by *Cirsium arvense* (Tulloch and Hoffman, 1982), whereas lupeyl acetates are found in *Cirsium arvense*, *Hieracium pilosella*, *Picris echioides* and *Sonchus oleraceus* (Tulloch and Hoffman, 1982; Jakupovic et al., 2011).

Most of the triterpenyl acetate distribution in our soil sample can thus be explained by Asteraceae species found in the surrounding vegetation, except for taraxeryl, glutinyl, multiflorenyl and δ -amyrin acetates (Table 2). Because current knowledge of the distribution of these compounds does not allow explanation of their presence in our sample, alternative explanations can be invoked, each exemplified by one or two compounds:

(i) Taraxeryl, glutinyl, δ -amyrin and multiflorenyl acetates may be produced by other Asteraceae spp. that occur in the catchment but have not yet benefitted from a complete phytochemical survey. Due to their belonging to genera in which other species are known to synthesize triterpenyl acetates, they are likely to do so as well (Table 2). In addition to *Cirsium arvense*, six *Cirsium* spp. have been described in the Lake Aydat catchment (*C. acaule*, *C. dissectum*, *C. eriophorum*, *C. erisithales*, *C. palustre* and *C. vulgare*; Antonetti et al., 2006), but whose pentacyclic triterpenyl acetate content has not been investigated. Because multiflorenyl acetate was described in *Cirsium* sp. and *C. benedictus* (Asteraceae; Ulubelen and Berkan, 1977; Kataria, 1995), it is tempting to suppose that one of the *Cirsium* spp. in the catchment could also produce multiflorenyl acetate. Similarly, because taraxeryl acetate was described in *Artemisia dalailamae*

(Yang et al., 1994), it could possibly originate from *Artemisia verlotiorum* or *A. vulgaris* (Asteraceae), both reported in the area. Likewise, δ -amyrin acetate was described in *Inula britannica*, while another species of the genera, *I. conyza*, grows in the area and could therefore be the source of this compound in the soil sample.

(ii) These compounds are produced by plants other than Asteraceae in the catchment. If the two preceding examples can help clarifying the presence of taraxeryl, δ -amyrin and multiflorenyl acetates, they cannot account for the presence of glutinyl acetate.

According to phytochemical data, this compound is only described in *Erythrophleum fordii* (Fabaceae: Tsao et al., 2008), in *Kalanchoe daigremontiana* (Crassulaceae: van Maarseveen and Jetter, 2009), in *Acer mandshuricum* (Aceraceae: Ding et al., 2010), in *Dorstenia* sp. and in *Maquira coriacea* (Moraceae: Woldu et al., 1988; Mitaine-Offer et al., 2001), none of these species or genus being found in the catchment except for *Acer* sp. (*A. pseudoplatanus*, *A. campestre* subsp. *campestre* and *A. platanoides*; Antonetti et al., 2006). In the area, the occurrence of *Acer* species could thus explain the presence of glutinyl acetate in the soil sample.

(iii) The compounds are produced by one or several of the already identified potential sources of triterpenyl acetates. Asteraceae spp. developed around Lake Aydat could synthesize distinct triterpenyl acetates distinct from those in the same species cited in the literature. Such a variability in chemodemes (i.e. biochemical races - plants from the same species producing distinct chemicals depending on location, environmental conditions or slight genetic differences) have, for example, been described for pentacyclic triterpene methyl ethers in New Zealand (Connor and Purdie, 1976).

(iv) Pentacyclic triterpenes are reputed to suffer structural rearrangements and double bond migration during diagenesis that could affect the original diversity of compounds (e.g. Rullkötter et al., 1994). This phenomenon that is well documented from laboratory experiments for compounds the oleanane series (Courtney et al., 1958; Coates et al., 1967), is also observed in sedimentary records (Rullkötter et al., 1994). Precisely, friedel-3-ene, glutin-5-ene, multiflor-7-ene and taraxer-14-ene are progressively rearranged to olean-18-ene, olean-12-ene and finally olean-13(18)-ene isomers. Such rearrangements have also been reported in hopane and fernane series (Nishimoto et al., 1968) in rock and sediment samples (Volkman et al., 1986; Paull et al., 1998). To our knowledge, the only studies about such reactions on triterpenyl acetates that were realised in the laboratory (Chatterjee et al.; 1976), showed the acid-induced transformation of taraxeryl acetate into β -amyrin acetate, which could further be transformed into δ -amyrin acetate. To our knowledge, the stability of pichierenyl or swertenyl acetates in natural environments remains totally to be checked. Thus, in the absence of available evidence, the possibility of rearrangements of triterpenyl acetates in soils can be excluded, further work being needed to ascertain whether this conclusion could be extended to sediments and especially over long durations.

4.3. Potential palaeoenvironmental implications

Few studies have reported the occurrence of triterpenyl acetates in soil and sediments (Logan et al., 1995; van Bergen et al., 1997; Xu et al., 2008; Oyo-Ita et al., 2010; Trendel et al., 2010). According to Oyo-Ita et al. (2010), this lack can be explained by the procedures generally applied, saponification entailing the loss of the acetate group. These authors also underline the possibility of natural hydrolysis of the

ester bond during riverine transport. They therefore attribute the presence of triterpenyl acetates in sediments to a local source. Despite the few reports, no origin was proposed for the compounds, probably because, except for lupeyl acetates, those detected (taraxeryl, germanicyl, δ -amyrin, β -amyrin acetates) were not of direct taxonomical interest. To our knowledge, our work constitutes the first report on bauerane, taraxastane, swertane and pichierane acetates in soil, which can be unequivocally linked to Asteraceae.

Asteraceae are defined as meadow and pasture species (semi-open to open habitats) that colonize disrupted environments and are able to conquer fallow land developed on abandoned arable land (Cronquist, 1980; Bouby and Billaud, 2001, 2005; Antonetti et al., 2006). Asteraceae pollen is frequently used as an anthropogenic indicator because it is found mainly in settlements next to cultivated fields, in mowed or grazing places, or in ruderal communities (i.e. growing in rubbles; Hicks, 1992; Court-Picon et al., 2006). Thus, the detection of triterpenyl acetates in soil and sediments could attest to both open vegetation and major perturbation in the catchment, under natural but more obviously anthropogenic influence.

The compounds could constitute diagnostic tracers of agriculture and other human activities. For example, the presence of various Asteraceae seeds in a Late Bronze Age (905-869 BC) settlement on the shore of Lake le Bourget was related to their ethnobotanical properties, their consumption as human food or livestock fodder, dye and/or medicine (Bouby and Billaud, 2001; 2005). Triterpenyl acetates thus extend the panel of biomarkers for past flora that can attest, in favourable cases, to human activities such as miliacin for millet (Jacob et al., 2008), other pentacyclic triterpene methyl ethers for Gramineae (Ohmoto et al., 1970; Jacob et al., 2005; Zocatelli et al.,

2010), *iso*- and *anteiso*- monomethyl alkanes for culinary and aromatic herbs from the Lamiaceae family (Huang et al., 2011) and more generally those reported by Evershed (2008) with respect to archaeology. As with these other biomarkers, triterpenyl acetates can constitute reliable tools for tracing the presence of plants of economic interest. They could reinforce other widely used techniques that allow detection of the presence of Asteraceae in any sedimentary or archaeological context: for example, for palynological studies that suffer from the under representation of Asteraceae in pollen records because of their dispersal mode (Andrieu et al., 1997).

5. Conclusions

Neutral lipids extracted from soils developed under pasture and meadow exhibit a wide array of pentacyclic triterpene structures bearing an acetate group at C-3. Among these, C-3 acetates with bauerane, taraxastane, swertane and pichierane skeletons are reported in natural archives for the first time. An exhaustive review of the phytochemical literature allowed us to unequivocally attribute some of the triterpenyl acetates to a single vegetation source within the Asteraceae family. This specificity confers to these compounds a high potential as tracers of Asteraceae – and thus of open environments - for palaeoenvironmental studies.

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Table captions

Table 1

Inventory of triterpenyl acetates in soil samples with peak number, retention times and mass spectral data.

Peak	Ret. time (t_R min)	Name	Trivial name	M^+	Significant MS fragments (decreasing abundance)
1	44.90	Tarax-14-en-3 β -yl acetate	Taraxeryl acetate	468	204, 189, 269, 218, 329, 344, 393, 453, 468
2	45.15	Olean-13(18)-en-3 β -yl acetate	δ -Amyrin acetate	468	189, 205, 218, 204, 203
3	45.28	Olean-12-en-3 β -yl acetate	β -Amyrin acetate	468	203, 218, 189, 257, 323, 393, 408, 453, 468
4	45.57	Bauer-8-en-3 β -yl acetate	Isobauerenyl acetate	468	229, 289, 241, 257, 341, 393, 409
5	46.23	Glutin-5-en-3 β -yl acetate	Glutinylnyl acetate	468	259, 274, 393, 408, 453, 468
6	46.25	Urs-12-en-3 β -yl acetate	α -Amyrin acetate	468	218, 189, 203, 257, 393, 408, 468, 453
7	46.28	Lup-20(29)-en-3 β -yl acetate	Lupeyl acetate	468	189, 204, 218, 297, 355, 453, 468, 393
8	47.62	Multiflor-7-en-3 β -yl acetate	Multiflorenyl acetate	468	205, 229, 241, 301, 262, 289
9	48.08	Bauer-7-en-3 β -yl acetate	Bauerenyl acetate	468	229, 289, 241, 393, 453, 468
10	48.40	Taraxast-20-en-3 β -yl acetate	ψ -Taraxasteryl acetate	468	189, 204, 249, 408, 468, 393
11	48.70	Taraxast-20(30)-en-3 β -yl acetate	Taraxasteryl acetate	468	189, 218, 204, 232, 249, 408, 262, 468
12	49.11	Pichier-8-en-3 β -yl acetate	Isopichierenyl acetate	468	289, 229, 241, 301, 393, 453, 468
13	49.73	Pichier-9(11)-en-3 β -yl acetate	Pichierenyl acetate	468	289, 301, 241, 229
14	49.86	Gammacer-16-en-3 β -yl acetate	Gammacerenyl acetate	468	187, 191, 327, 408, 468
15	51.63	Pichier-7-en-3 β -yl acetate	Swertenyl acetate	468	289, 229, 241, 301, 393, 453, 468

Table 2

Known occurrences of triterpenyl acetates in soil sample from Lake Aydat

Catchment.^{a,b}

	Taraxeryl acetate (1)	6-Amyrin acetate (2)	β-Amyrin acetate (3)	Isobauerenyl acetate (4)	Glutinyl acetate (5)	α-Amyrin acetate (6)	Lupeyl acetate (7)	Multiflorenyl acetate (8)	Bauerenyl acetate (9)	ψ-Taraxasteryl acetate (10)	Taraxasteryl acetate (11)	Isopichierenyl acetate (12)	Pichierenyl acetate (13)	Gammacerenyl acetate (14)	Swertenyl acetate (15)	References
(a) Asteraceae species																
<i>Arctium lappa</i> L.											X					Jakupovic et al., 2011
<i>Carduus nutans</i>											X					Jordon-Thaden and Louda, 2003
<i>Cirsium arvense</i>		X				X	X			X	X					Dutta and Ray, 1972; Tulloch and Hoffman, 1982; Jakupovic et al., 2011
<i>Hieracium pilosella</i> L.						X										Jakupovic et al., 2011
<i>Onopordum acanthium</i>										X						Khalilova et al., 2003, 2004; Shoeb et al., 2007
<i>Picris echioides</i>							X									Jakupovic et al., 2011
<i>Picris hieracioides</i>		X	X			X	X		X	X	X	X	X	X	X	Shiojima et al., 1989a; b; 1995; Jakupovic et al., 2011
<i>Solidago virga-aurea</i>		X														Choi et al., 2004
<i>Sonchus oleraceus</i> L.						X	X				X					Jakupovic et al., 2011
<i>Taraxacum officinale</i>											X					Hänsel et al., 1980
Total (a)		X	X			X	X		X	X	X	X	X	X	X	
(b) Asteraceae genera																
<i>Artemisia</i> (2)	X	X				X	X		X		X					Lao et al., 1983; 1984; Yang et al., 1994; Jakupovic et al., 2011
<i>Cirsium</i> (7)		X				X	X	X		X	X					Dutta and Ray, 1972; Ulubelen and Berkan, 1977; Tulloch and Hoffman, 1982; Jakupovic et al., 2011
<i>Inula</i> (1)		X				X				X						Ahmad and Ismail, 1991; Jakupovic et al., 2011
Total (a)+(b)	X	X	X	X		X	X	X	X	X	X	X	X	X	X	
(c) Other taxa																
<i>Acer</i>		X		X												van Bergen et al., 1997; Ding et al., 2010
<i>Euphorbia</i>	X	X	X	X		X	X	X		X						Dutta and Ray, 1972; Ahmad and Fizza, 1986; Liu et al., 2001; Shi et al., 2005; Haba et al., 2007
Total (a)+(b)+(c)	X	X	X	X	X	X	X	X	X	X	X	X	X	X	X	

^a (a) Asteraceae species in Lake Aydat catchment; (b) Asteraceae genera in which triterpenyl acetates have been described; ^b numbers into parentheses refer to number of species of a given genera described in the catchment according to Antonetti et al. (2006).

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Figure captions

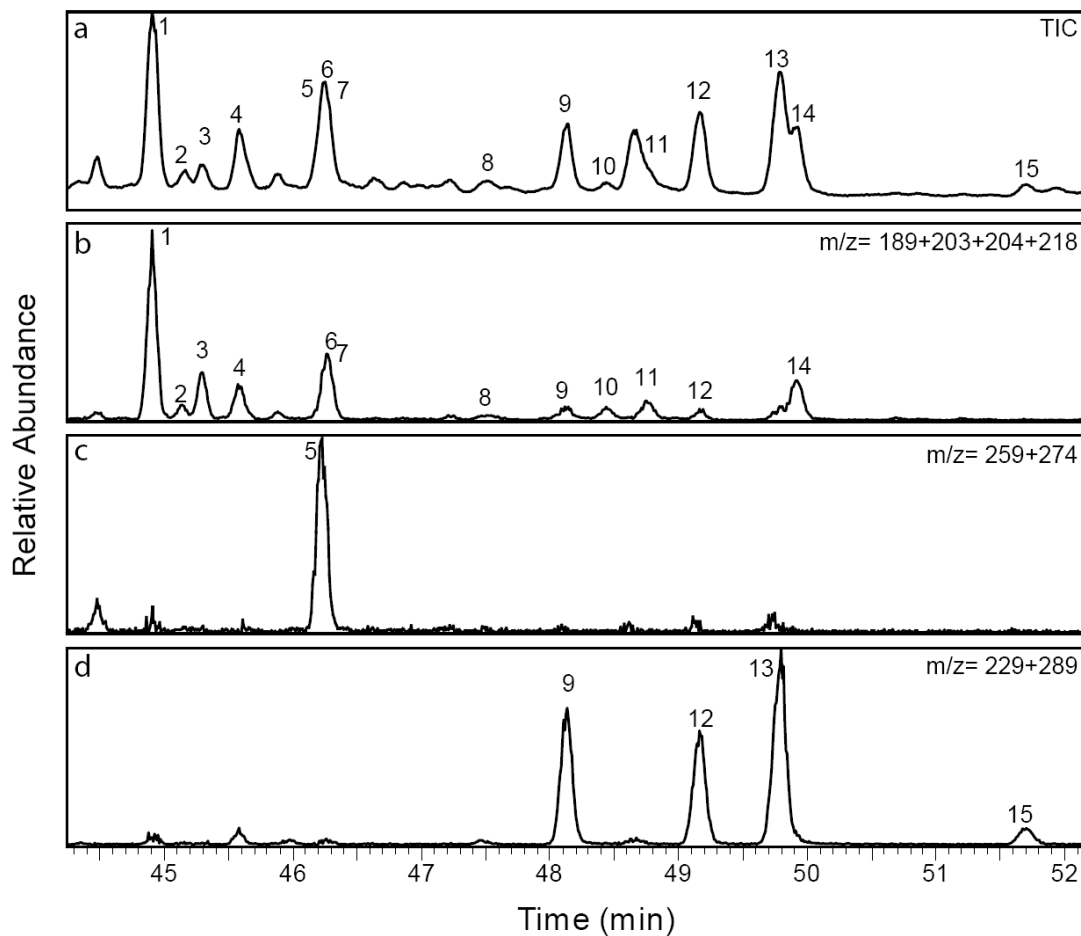


Fig. 1. Example of distribution of triterpenyl acetates in ketone/acetate fraction from a soil sample from catchment of Lake Aydat, under pasture. a TIC); b m/z 189+203+204+218 chromatogram; c m/z 259+274 chromatogram; d m/z 229+289 chromatogram. The MS and t_R data and assignment of compounds 1 to 15 are given in Table 1. The spectra are provided in Fig. 2.

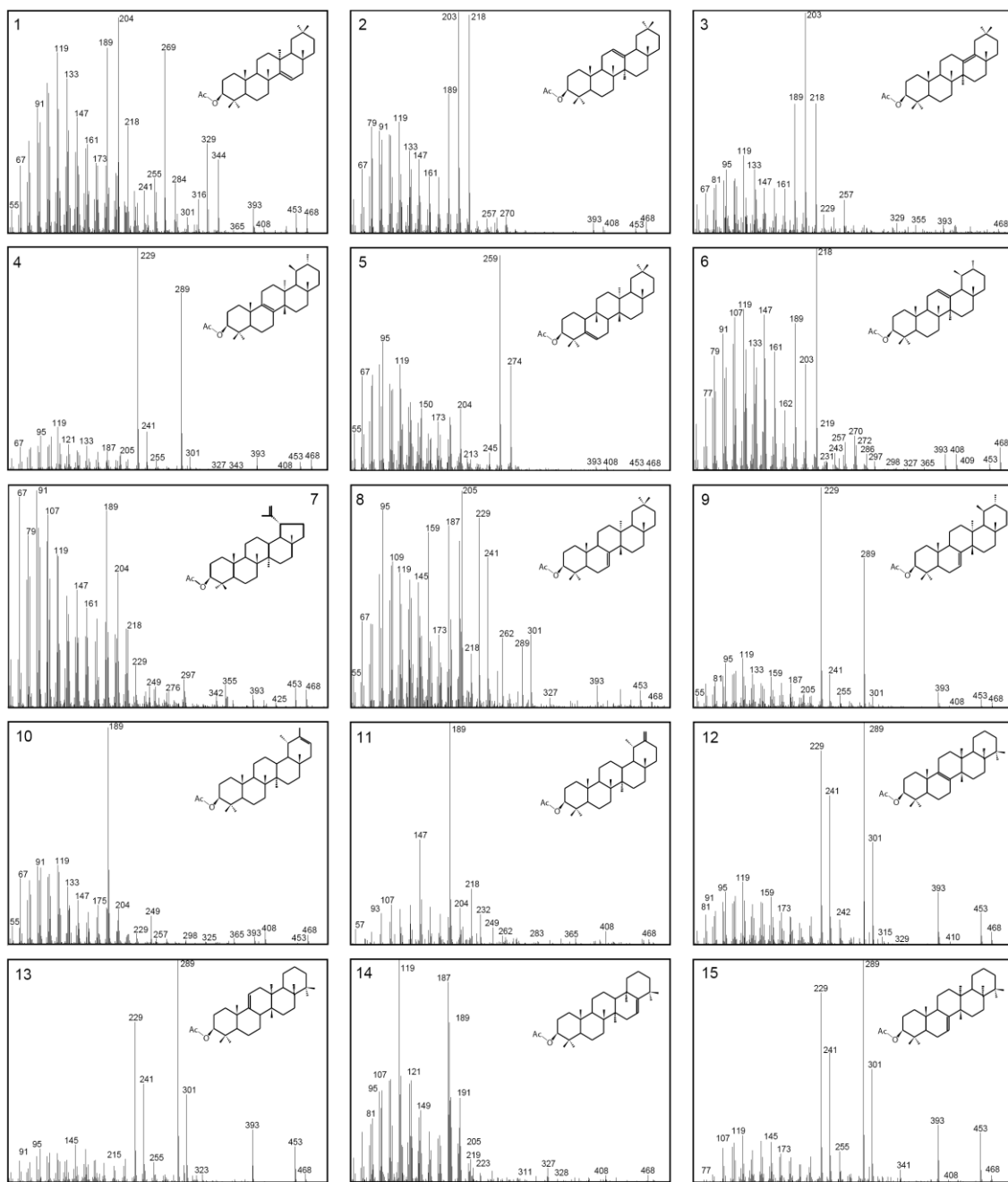


Fig. 2. Mass spectra and structure of compounds in soil sample. Numbers refer to Table 1 and Fig. 1. Mass spectra of glutinyl, α -amyrin and lupeyl acetates were recorded for the standard compound because of GC co-elution with the soil sample.